BACKGROUND: IGF2BP2 is an RNA binding protein that post-transcriptionally regulates IGF2 during developmental stages. IGF2 mRNA-binding protein family (IGF2BP1, 2 and 3) bind UTR region of IGF2 transcripts and are implicated to in transport of RNA targets to enable protein synthesis at specific locations in the cytoplasm. IGF2BP family members show tissue-selective expression during development. IGF2BP2 is expressed in fetal lung, kidney, thymus, placenta and liver. Genetic valuation in IGF2BP2 was implicated to interact with Fetal Malnutrition on glucose metabolism.

RIP-CERTIFIED ANTIBODY:
Posttranscriptional regulation of gene expression is a ribonucleoprotein-driven process, which involves RNA binding proteins (RBPs) and non-coding RNAs that affect splicing, nuclear export, subcellular localization, mRNA decay and translation. The RNP Immunoprecipitation-Chip (RIP-Chip), RIP-Seq and RIP-RTPCR allow the identification of multiple RNA targets of RBPs globally and within the context of a cell extract. Antibodies specific to the RNA binding protein of interest are used to co-immunoprecipitate the RNA binding protein and the associated subset of mRNAs. The mRNA content is interrogated using standard microarray or sequencing technology. RIP-Certified Antibody is validated for use in RNP Immunoprecipitation (RIP) in conjunction with the RIP-Assay Kit distributed from MBL. Its ability to immunoprecipitate mRNAs and RBPs complex was confirmed by quantitative and qualitative analysis on NanoDrop, Bioanalyzer and RT-PCR or microarray.

SOURCE: This antibody was purified from rabbit serum by affinity column chromatography. The rabbit was immunized with KLH conjugated synthetic peptide, KQEQEKYPQGVASQSK corresponding to 583-599 aa.

FORMULATION: 200 µL volume of PBS containing 50% glycerol, pH 7.2. No preservative is contained.

STORAGE: This antibody solution is stable for one year from the date of purchase when stored at -20°C.

INTENDED USE:
For Research Use Only. Not for use in diagnostic procedures.

REACTIVITY: This antibody reacts with human IGF2BP2 (~66 kDa) on Western blotting, Immunoprecipitation and RNP Immunoprecipitation.

REFERENCES:
3) van Hoek, M. et al., Diabetes E publication, Mar 3, (2009)

SPECIES CROSS REACTIVITY:

<table>
<thead>
<tr>
<th>Species</th>
<th>Human</th>
<th>Mouse</th>
<th>Rat</th>
<th>Hamster</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cells</td>
<td>K562, 293T, HeLa</td>
<td>NIH/3T3</td>
<td>Rattl</td>
<td>CHE</td>
</tr>
<tr>
<td>Reactivity on WB</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
</tbody>
</table>
The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

PROTOCOLS:

RNP Immunoprecipitation

Some buffer and reagents are included in the RIP-Assay Kit (code: RN1001). Please also refer to the protocol packaged in the RIP-Assay Kit.

**[Material Preparation]**

1. **Lysis Buffer (+)**
   Before using the Lysis Buffer, protease inhibitors, RNase inhibitors, and DTT are added to the Lysis Buffer at the appropriate concentration.

2. **Wash Buffer (+)**
   Before using the Wash Buffer, DTT is added to the Wash Buffer at the appropriate concentration.

**Protocol**

1) Wash 1.2 x 10^7 cells 2 times with PBS and resuspend them with 500 µL of ice-cold Lysis Buffer (+) containing appropriate protease inhibitors, RNase inhibitors, and DTT. Vortex for 10 seconds. Leave on ice for 10 minutes.

2) Centrifuge the tube at 12,000 x g for 5 minutes at 4°C and transfer the supernatant to another tube.

3) Add 25 µL of 50% protein A agarose beads slurry suspended in Lysis Buffer (+) into the supernatant. Incubate it at 4°C with rotating for 1 hour.

4) Centrifuge the tube at 2,000 x g for 1 minute at 4°C and transfer the supernatant to another tube (precleared sample).

5) Mix both 25 µL of 50% protein A agarose beads slurry suspended in nuclease-free PBS and Normal Rabbit IgG (RIP-Assay Kit) or Anti-IGF2BP2 (IMP2) pAb (RN008P) at the amount of suggested in the APPLICATIONS, and then add 1 mL of Wash buffer (+) into each tube. Incubate with gentle agitation for 1 hour at 4°C.

6) Wash the beads once with ice-cold Lysis Buffer (+) (centrifuge the tube at 2,000 x g for 1 minute). Carefully discard the supernatant using a pipettor without disturbing the beads.

7) Add 500 µL of cell lysate (precleared sample of step 4), then incubate with gentle agitation for 3 hours at 4°C.

8) Wash the beads 4 times with Wash Buffer (+) (centrifuge the tube at 2,000 x g for 1 minute).

9) Add 400 µL of Master mix solution (Solution I: Solution II = 10 µL: 390 µL). Vortex for 10 seconds.

10) Add 250 µL of Solution III. Vortex for 10 seconds.

11) Centrifuge the tube at 2,000 x g for 2 minutes.

12) Transfer the supernatant to the tube containing 2 µL of Solution IV.

13) Add 600 µL of ice-cold 2-propanol and place at -20°C for 20 minutes. Centrifuge the tube at 12,000 x g for 10 minutes.

14) Wash the pellet 2 times with 0.5 mL of ice-cold 70% Ethanol and dry up the pellet for 5-15 minutes.

15) Dissolve the pellets in nuclease-free water.

16) RNA was quantified with NanoDrop (Thermo Fisher Scientific Inc.) and the RNA quality was analyzed with Bioanalyzer (Agilent Technologies, Inc.).

**Analysis of isolated RNA with Bioanalyzer.**

<table>
<thead>
<tr>
<th>Average of the RNA Quantity (n=2)</th>
<th>Antibody</th>
<th>RNA (ng)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal Rabbit IgG</td>
<td>73.0</td>
<td></td>
</tr>
<tr>
<td>anti-IGF2BP2</td>
<td>756.0</td>
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</tr>
<tr>
<td>Total RNA</td>
<td>220966.7</td>
<td></td>
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</tbody>
</table>

(Positive control for RNP Immunoprecipitation; K562)

**SDS-PAGE & Western blotting**

1) Wash 1 x 10^7 cells 3 times with PBS and suspend them in 1 mL of Laemmli’s sample buffer.

2) Boil the samples for 2 minutes and centrifuge. Load 10 µL of the sample per lane in a 1 mm thick SDS-polyacrylamide gel for electrophoresis.

**Western blotting analysis of IGF2BP2 expression in K562 (1), 293T (2), HeLa (3), NIH/3T3 (4) and Rat1 (5) using RN008P.**
3) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm² for 1 hour in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% Methanol). See the manufacturer's manual for precise transfer procedure.

4) To reduce nonspecific binding, soak the membrane in 10% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature, or overnight at 4°C.

5) Incubate the membrane with primary antibody diluted with PBS, pH 7.2 containing 1% skimmed milk as suggested in the APPLICATIONS for 1 hour at room temperature. (The concentration of antibody will depend on condition.)

6) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 minutes x 3 times).

7) Incubate the membrane with the 1:10,000 Anti-IgG (Rabbit) pAb-HRP (MBL; code no. 458) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature.

8) Wash the membrane with PBS-T (5 minutes x 3 times).

9) Wipe excess buffer on the membrane, then incubate it with appropriate chemiluminescence reagent for 1 minute. Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.

10) Expose to an X-ray film in a dark room for 1 minute. Develop the film as usual. The condition for exposure and development may vary.

(Positive controls for Western blotting; K562, 293T, HeLa, NIH/3T3, Rat1)

Immunoprecipitation

1) Wash 1 x 10⁷ cells 2 times with PBS and resuspend them with 1 mL of ice-cold Lysis buffer (RIP-Assay Kit) containing appropriate protease inhibitors, then sonicate briefly (up to 10 seconds).

2) Centrifuge the tube at 12,000 x g for 5 minutes at 4°C and transfer the supernatant to another tube.

3) Add 20 µL of 50% protein A agarose beads slurry resuspended in nuclease-free PBS and Normal Rabbit IgG (RIP-Assay Kit) or Anti-IGF2BP2 (IMP2) pAb (RN008P) at the amount of suggested in the APPLICATIONS, and then add 1 mL of Wash buffer into each tube. Incubate with gentle agitation for 1 hour at 4°C.

4) Wash the beads once with ice-cold Lysis Buffer (centrifuge the tube at 2,000 x g for 1 minute). Carefully discard the supernatant using a pipettor without disturbing the beads.

5) Add 250 µL of cell lysate (precleared sample of step 4), then incubate with gentle agitation for 1 hour at 4°C.

6) Wash the beads 4 times with Wash Buffer (centrifuge the tube at 2,000 x g for 1 minute).

7) Resuspend the beads in 20 µL of Laemmli’s sample buffer, boil for 3-5 minutes, and centrifuge for 5 minutes. Use 20 µL/lane for the SDS-PAGE analysis.

(See SDS-PAGE & Western blotting.)

(Positive control for Immunoprecipitation; K562)

RELATED PRODUCTS:
Please visit our website at https://ruo.mbl.co.jp/je/rip-assay/.