POLYCLONAL ANTIBODY

Anti-LC3 pAb

<table>
<thead>
<tr>
<th>Code No.</th>
<th>Quantity</th>
<th>Form</th>
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<tbody>
<tr>
<td>PM036</td>
<td>100 µL</td>
<td>Purified IgG</td>
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</table>

BACKGROUND: Macropathway mediates the bulk degradation of cytoplasmic components. These components are delivered to lysosomes via autophagosomes. The microtubule-associated protein 1 light chain 3 (LC3), a homologue of yeast Atg8 (Avt7/Apg8), localizes to autophagosomal membranes after post-translational modifications. The C-terminal fragment of LC3 is cleaved immediately following synthesis to yield a cytosolic form called LC3-I. A subpopulation of LC3-I is further converted to an autophagosome-associating form, LC3-II. This antibody can detect both forms of LC3.

SOURCE: This antibody was purified from rabbit serum using protein A agarose. The rabbit was immunized with the recombinant human LC3 [MAP1LC3B (1-120 aa)].

FORMULATION: 100 µL volume of PBS containing 50% glycerol, pH 7.2. No preservative is contained.

STORAGE: This antibody solution is stable for one year from the date of purchase when stored at -20°C.

REACTIVITY: This antibody reacts with LC3 (MAP1LC3A, B, C) on Western blotting, Immunoprecipitation, Immunocytochemistry, and Flow cytomtry. It does not react with GABARAP and GATE-16.

APPLICATIONS:
- Western blots: 1:1,000 for chemiluminescence detection system
- Immunoprecipitation: 2 µL/300 µL of cell extract from 1 x 10^7 cells
- Immunohistochemistry: 1:1,000-1:2,000
  - Heat treatment is necessary for paraffin embedded sections.
  - Microwave oven; 2 times for 10 minutes each in 10 mM citrate buffer (pH 6.3)
- Immunocytochemistry: 1:500-1:1,000
- Flow cytometry: 1:200 (final concentration)
- Immuno-electron microscopy: Not tested*
- *It is reported that this antibody can be used in this application in the reference number 4).

Detailed procedure is provided in the following PROTOCOLS.


SPECIES CROSS REACTIVITY:

<table>
<thead>
<tr>
<th>Species</th>
<th>Human</th>
<th>Mouse</th>
<th>Rat</th>
<th>Hamster</th>
<th>Other*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cells</td>
<td>HeLa, 293T, Raji</td>
<td>NIH/3T3, MEF, PC12, Rat1</td>
<td>CHO</td>
<td>Not tested</td>
<td></td>
</tr>
</tbody>
</table>

Reactivity on WB + + + +

*It is reported that this antibody reacts zebrafish tissues in the reference number 1) and 3).

REFERENCES:
2) Wang, Y., et al., Cell 171, 331-345.e22 (2017) [IC]
4) Fujita, N., et al., Elife 6, e23367 (2017) [Immu-No-EM]
6) Kaminsky, V. O., et al., Autophagy 8, 1032-1044 (2012) [IC]

As this antibody is widely used, many researches have been reported. These references are a part of such reports.

Western blot analysis of LC3 in positive control (PM036-PN) (1), NIH/3T3 (2), PC12 (3), Rat1 (4), CHO (5), HeLa (6), 293T (7), Raji (8) and mouse brain lysate (9) using PM036.
Western blot analysis of LC3 in Atg5−/− MEF (left) and wild-type MEF (right) using PM036.

Atg5−/− MEF was kindly provided by Dr. Noboru Mizushima, M.D., Ph.D. (Department of Biochemistry and Molecular Biology, Graduate School and Faculty of Medicine, The University of Tokyo)

**PROTOCOLS:**

**SDS-PAGE & Western Blotting**

1) Wash 1 x 10^7 cells 3 times with PBS and suspend them in 1 mL of Laemmli's sample buffer, then sonicate briefly (up to 20 sec.).

2) Boil the samples for 3 minutes and centrifuge. Load 10 μL of sample per lane on a 1-mm-thick SDS-polyacrylamide gel (15% acrylamide) and carry out electrophoresis.

3) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm² for 1 hour in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% MeOH). See the manufacturer's manual for precise transfer procedure.

4) To reduce nonspecific binding, place the membrane in 10% skimmed milk (in PBS, pH 7.2) overnight at 4°C.

5) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 minutes x 3 times).

6) Incubate the membrane with primary antibody diluted with 1% skimmed milk (in PBS, pH 7.2) as suggested in the APPLICATIONS for 1 hour at room temperature. (The concentration of antibody will depend on the condition.)

7) Wash the membrane with PBS-T (5 minutes x 3 times).

8) Incubate the membrane with 1:10,000 Anti-IgG (Rabbit) pAb-HRP (MBL; code no. 458) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature.

9) Wash the membrane with PBS-T (5 minutes x 3 times).

10) Wipe excess buffer on the membrane, then incubate it with appropriate chemiluminescence reagent for 1 minute.

11) Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.

12) Expose to an X-ray film in a dark room for 3 minutes.

13) Develop the film as usual. The condition for exposure and development may vary.

(Positive controls for Western blotting: PM036-PN, NIH/3T3, PC12, Rat1, CHO, HeLa, HEK293T, Raji and Mouse brain lysate)

**Immunocytochemistry**

1) Spread the cells in the nutrient condition on a glass slide, then incubate in a CO₂ incubator for one night.

2) Remove the culture supernatant by careful aspiration.

3) To obtain serum-starved conditions, culture the cells with Hank’s solution or DMEM for 2-4 hours at 37°C.

4) Fix the cells by immersing them in 4% paraformaldehyde (PFA)/PBS for 10 minutes at room temperature (20~25°C).

5) Wash the slide 2 times with PBS.

6) Immerse the slides in 100 μg/mL of Digitonin for 10 minutes at room temperature.

7) Wash the slides 2 times with PBS.

8) Add the primary antibody diluted with PBS as suggested in the APPLICATIONS onto the cells and incubate for 1 hour at room temperature. (Optimization of antibody concentration or incubation condition is recommended if necessary.)

9) Wash the slides 2 times with PBS.

10) Add 200 μL of 1:500 Alexa Fluor® 488 Goat Anti-rabbit IgG (Invitrogen; code no. A11008) diluted with PBS onto the cells. Incubate for 30 minutes at room temperature. Keep out light by aluminum foil.

11) Wash the slides 2 times with PBS.

12) Counterstain with DAPI for 5 minutes at room temperature.

13) Wash the slides 2 times with PBS.

14) Wipe excess liquid off the slide but take care not to touch the cells. Never leave the cells to dry.

15) Promptly add mounting medium onto the slide, then put a cover slip on it.

(Positive control for Immunocytochemistry; NRK)
**Immunoprecipitation of LC3 from HeLa (1, 2) and NIH/3T3 (3, 4) with Rabbit IgG (1, 3) or PM036 (2, 4).** After immunoprecipitated with the antibody, immunocomplex was resolved on SDS-PAGE and Immunoblotted with M186-3.

**Immunoprecipitation**

1) Wash the cells 3 times with PBS and suspend with 10 volume of cold Lysis buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 1% NP-40] containing appropriate protease inhibitors. Incubate it at 4°C with rotating for 30 minutes, then sonicate briefly (up to 10 seconds).
2) Centrifuge the tube at 12,000 x g for 10 minutes at 4°C and transfer the supernatant to another tube.
3) Mix 20 μL of 50% protein A agarose beads slurry resuspended in 300 μL of IP buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 0.05% NP-40] with primary antibody as suggested in the APPLICATIONS. Incubate with gentle agitation for 1 hour at 4°C.
4) Wash the beads 1 time with 1 mL of IP buffer.
5) Add 300 μL of cell lysate (prepared sample from step 2) into the tube. Incubate with gentle agitation for 1 hour at 4°C.
6) Wash the beads 4 times with 1 mL of IP buffer.
7) Resuspend the beads in 20 μL of Laemmli’s sample buffer, boil for 3 minutes, and centrifuge for 5 minutes.
8) Load 10 μL of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel (15% acrylamide) for electrophoresis.
9) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm² for 1 hour in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% Methanol). See the manufacture's manual for precise transfer procedure.
10) To reduce nonspecific binding, soak the membrane in 10% skimmed milk (in PBS, pH 7.2) overnight at 4°C.
11) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 minutes x 3 times).
12) Incubate the membrane with 1 μg/mL of Anti-LC3 mAb (MBL; code no. M186-3) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature. (The concentration of antibody will depend on the conditions.)
13) Wash the membrane with PBS-T (5 minutes x 3 times).
14) Incubate the membrane with 1:10,000 Anti-IgG (Mouse) pAb-HRP (MBL; code no. 330) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature.
15) Wash the membrane with PBS-T (5 minutes x 3 times).
16) Wipe excess buffer on the membrane, then incubate it with appropriate chemiluminescence reagent for 1 minute.
17) Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.
18) Expose to an X-ray film in a dark room for 3 minutes.
19) Develop the film as usual. The condition for exposure and development may vary.

(Positive controls for Immunoprecipitation; HeLa and NIH/3T3)

**Immunohistochemical staining for paraffin-embedded sections**

1) Deparaffinize the sections with Xylene 3 times for 3-5 minutes each.
2) Wash the slides with Ethanol 3 times for 3-5 minutes each.
3) Wash the slides with PBS 3 times for 3-5 minutes each.
4) Heat treatment
   Heat treatment by Microwave:
   Place the slides put on staining basket in 500 mL beaker with 500 mL of 10 mM citrate buffer (pH 6.3). Cover the beaker with plastic wrap, then process the slides 2 times for 10 minutes each at 500 W with microwave oven. Let the slides cool down in the beaker at room temperature for about 40 minutes.
5) Remove the slides from the citrate buffer and cover each section with 3% H₂O₂ for 10 minutes at room temperature to block endogenous peroxidase activity. Wash 3 times in PBS for 5 minutes each.
6) Remove the slides from PBS, wipe gently around each section and cover tissues with blocking buffer (20 mM HEPES, 1% BSA, 135 mM NaCl) for 5 minutes to block non-specific staining. Do not wash.
7) Tip off the blocking buffer, wipe gently around each section and cover tissues with primary antibody diluted with blocking buffer as suggested in the APPLICATIONS.
8) Incubate the sections for 1 hour at room temperature.
9) Wash the slides 3 times in PBS for 5 minutes each.
10) Wipe gently around each section and cover tissues with Histostar™ (rabbit) (MBL; code no. 8466). Incubate for 1 hour at room temperature. Wash as in step 9).
11) Visualize by reacting for 5 minutes with Histostar™ DAB Substrate Solution (MBL; code no. 8469). *DAB is a suspect carcinogen and must be handled with care. Always wear gloves.
12) Wash the slides in water for 5 minutes.
13) Counter stain in hematoxylin for 1 minute, wash the slides 3 times in water for 5 minutes each, and then immerse the slides in PBS for 5 minutes. Dehydrate by immersing in Ethanol 3 times for 3 minutes each, followed by immersing in Xylene 3 times for 3 minutes each.
14) Now ready for mounting.

Flow cytometric analysis of MEF, NRK and Atg5⁻/⁻ MEF cells under starved or nutrient conditions using PM036. Fluorescence intensity of LC3 on starved cells was reduced. Atg5⁻/⁻ MEF was provided by Dr. Mizushima M.D. Ph.D.

Flow cytometric analysis for adherent cells
We usually use Fisher tubes or equivalents as reaction tubes for all steps after 4).
1) To obtain serum-starved conditions, culture the cells with Hank’s solution or DMEM for 4 hours at 37°C.
2) Detach the cells from culture dish by trypsinization. *Excessive trypsinization may reduce the antigenecity.
3) Wash the cells with PBS.
4) Fix the cells with 4% paraformaldehyde (PFA)/PBS for 15 minutes at room temperature (20-25°C). Wash the cells 2 times with PBS.
5) Permeabilize the cells with 100 μg/mL of Digitonin for 15 minutes at room temperature. Wash the cells 2 times with PBS.
6) Resuspend the cells with PBS (5x10⁶ cells/mL).
7) Add 50 μL of the cell suspension into each tube, and centrifuge at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
8) Add 40 μL of the primary antibody at the concentration as suggested in the APPLICATIONS diluted in the washing buffer. Mix well and incubate for 30 minutes at room temperature.
9) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
10) Add FITC-conjugated anti-rabbit IgG antibody diluted with the washing buffer. Mix well and incubate for 15 minutes at room temperature.
11) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
12) Resuspend the cells with 500 μL of the washing buffer and analyze by a flow cytometer.

RELATED PRODUCTS:
PM036 Anti-LC3 pAb [WB, IP, IC, IHC, FCM]
M152-3 Anti-LC3 mAb (4E12) [WB, IP, IC, FCM, EM]
M186-3 Anti-LC3 mAb (8E10) [WB]
PD014 Anti-LC3 pAb [WB]
M186-7 Anti-LC3 mAb-HRP-DirectT (8E10)
PM045 Anti-p62 (SQSTM1) pAb
M162-3 Anti-p62 (SQSTM1) (Human) mAb (5F2)
PM066 Anti-p62 C-terminal pAb
M217-3 Anti-Phospho-p62 (SQSTM1) (Ser351) mAb (5D5)
PM074 Anti-Phospho-p62 (SQSTM1) (Ser351) pAb
D343-3 Anti-Phospho-p62 (SQSTM1) (Ser403) mAb (4F6)
D344-3 Anti-Phospho-p62 (SQSTM1) (Ser403) mAb (4C8)
PD017 Anti-Beclin 1 pAb
PM037 Anti-GABARAP pAb
M135-3 Anti-GABARAP mAb (1F4)
PM038 Anti-GATE-16 pAb
PD041 Anti-Atg2A pAb
PM034 Anti-Atg3 p Ab
M134-3 Anti-Atg4B mAb (9H5)
PM050 Anti-Atg5 p Ab
PM039 Anti-Atg7 (Human) pAb
PD042 Anti-Atg9A pAb
M151-3 Anti-Atg10 (Human) mAb (5A7)
M154-3 Anti-Atg12 (Human) mAb (6E5)
PD036 Anti-Atg13 (Human) pAb
PD026 Anti-Atg14 pAb
PM040 Anti-Atg16L pAb
M160-3 Anti-UVRAG mAb (1H4)
PD027 Anti-Rubicon (Human) pAb
PM069 Anti-NRF2 pAb
M200-3 Anti-NRF2 mAb (1F2)
PD037 Anti-Tel2 pAb
PM072 Anti-VMF1 pAb
PM035 Normal Rabbit IgG
PM036-PN Positive control for anti-LC3 antibody

Other related antibodies and kits are also available. Please visit our web site at [http://ruo.mbl.co.jp](http://ruo.mbl.co.jp)