# For Research Use Only. Not for use in diagnostic procedures.



#### MONOCLONAL ANTIBODY

# Mouse IgG2b (isotype control)-PE

Code No. Clone Subclass Quantity Concentration M077-5 3D12 Mouse IgG2b  $\kappa$  1 mL (50 tests) 10  $\mu$ g/mL

**SOURCE:** This antibody was purified from hybridoma (clone 3D12) supernatant using protein A agarose. This hybridoma was established by fusion of mouse myeloma cell P3U1 with Balb/c mouse lymph nodes immunized with KLH.

**FORMULATION:** 10 μg IgG in 1 mL volume of PBS containing 1% BSA and 0.1% ProClin 150.

**STORAGE:** This antibody solution is stable for one year from the date of purchase when stored at 4°C.

**REACTIVITY:** No specific binding is detected on human peripheral blood leukocytes.

#### **APPLICATION:**

Flow cytometry; 20 µL (ready for use)

This antibody can be used as a negative isotypic control. The concentration will depend on condition.

Detailed procedure is provided in the following **PROTOCOLS**.

# **INTENDED USE:**

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The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

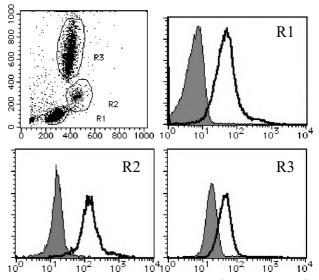
# **PROTOCOLS:**

# Flow cytometric analysis for floating cells

We usually use Fisher tubes or equivalents as reaction tubes for all steps described below.

- 1) Wash the cells 3 times with washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.09% NaN<sub>3</sub>].
  - \*Azide may react with copper or lead in plumbing system to form explosive metal azides. Therefore, always flush plenty of water when disposing materials containing azide into drain.
- 2) Resuspend the cells with washing buffer  $(5x10^6 \text{ cells/mL})$ .
- 3) Add 50 μL of the cell suspension into each tube, and centrifuge at 500 x g for 1 minute at room temperature (20~25°C). Remove supernatant by careful aspiration.
- 4) Add 20  $\mu L$  of Clear Back (human Fc receptor blocking reagent, MBL; code no. MTG-001) to the cell pellet after tapping. Mix well and incubate for 5 minutes at room temperature.

- 5) Add the primary antibody at the amount as suggested in the **APPLICATION**. Mix well and incubate for 30 minutes at room temperature.
- 6) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 7) Resuspend the cells with 500  $\mu L$  of the washing buffer and analyze by a flow cytometer.



Flow cytometric analysis of mouse IgG2b reactivity on lymphocyte (R1), monocyte (R2) and granulocyte (R3). Shaded histograms indicate the reaction of M077-5 to the cells. Open histograms indicate the reaction of PE labeled anti-HLA-A24 (K0209-5) to the cells.

# Flow cytometric analysis for whole blood cells

We usually use Falcon tubes or equivalents as reaction tubes for all steps described below.

- 1) Add the primary antibody at the amount as suggested in the **APPLICATION** into each tube.
- 2) Add 100  $\mu$ L of whole blood into each tube. Mix well and incubate for 30 minutes at room temperature (20~25°C).
- 3) Add 1 mL of the washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.09% NaN<sub>3</sub>] followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 4) Lyse with OptiLyse C (for analysis on Beckman Coulter instruments) or OptiLyse B (for analysis on BD instruments), using the procedure recommended in the

respective package inserts.

- 5) Add 1 mL of H<sub>2</sub>O to each tube and incubate for 10 minutes at room temperature.
- 6) Centrifuge at 500 x g for 1 minute at room temperature.
- 7) Add 1 mL of washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 8) Resuspend the cells with 500  $\mu L$  of the washing buffer and analyze by a flow cytometer.

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