

Smart-IP Series

Anti-Myc-tag mAb-Magnetic Beads

CODE No. M047-11
CLONALITY Monoclonal
CLONE PL14
ISOTYPE Mouse IgG1
QUANTITY 20 tests (Slurry: 1 mL)

SOURCE Purified IgG from mouse ascites fluid
IMMUNOGEN 6myc-tagged fusion protein
FORMULATION 15 mg magnetic beads in 1 mL PBS/0.1% BSA/0.09% NaN₃

*Azide may react with copper or lead in plumbing system to form explosive metal azides. Therefore, always flush plenty of water when disposing materials containing azide into drain.

STORAGE This beads suspension is stable for one year from the date of purchase when stored at 4°C.

APPLICATION-CONFIRMED

Immunoprecipitation 50 µL of beads slurry/sample

*The purification capacity of Anti-Myc-tag mAb-Magnetic Beads varies depending upon the characteristics of a Myc-tagged protein.
For example, 50 µL of beads slurry bounds 0.4 µg of a Myc-tagged protein (35 kDa).

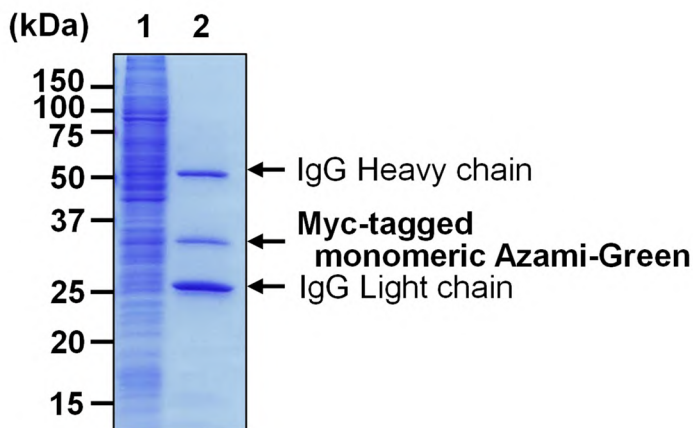
REFERENCE 1) Jain, P., *et al.*, *Oncogene* **36**, 6348-6358 (2017) [Co-IP]

For more information, please visit our website at <https://ruo.mbl.co.jp/>.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

Immunoprecipitation

- 1) Wash 1×10^7 cells 3 times with PBS and suspend with 1 mL of cold Lysis buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 1% NP-40] containing appropriate protease inhibitors.
- 2) Centrifuge the tube at 12,000 x g for 5 min. at 4°C and transfer the supernatant to another tube.
- 3) Add magnetic beads as suggested in the **APPLICATION** and purified Myc-tagged protein into 300 μ L of the supernatant prepared in step 2). Mix well and incubate with gentle agitation for 1 hr. at 4°C.
- 4) Place the tube on the magnetic rack (MBL, code no. 3190) for a few seconds.
- 5) Remove the supernatant.
- 6) Wash the beads 4 times with 1 mL of cold Wash buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 0.05% NP-40] (place the tube on the magnetic rack for a few seconds).
- 7) Resuspend the magnetic beads in 20 μ L of Laemmli's sample buffer, boil for 2 min., and place the tube on the magnetic rack for a few seconds.
- 8) Load 20 μ L of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel and carry out electrophoresis.
- 9) Visualize the protein bands by CBB staining.



Immunoprecipitation of Myc-tagged protein

Sample: 293T cell lysate from 3×10^6 cells + Myc-tagged monomeric Azami-Green 10 μ g

Lane 1: Input (5 μ L/lane)

Lane 2: Post-IP beads of Anti-Myc-tag mAb (MBL, code no. M047-11)