For Research Use Only. Not for use in diagnostic procedures.



MONOCLONAL ANTIBODY

Anti-Msx2

Code No. Clone Subclass Quantity Concentration M027-3 2E12 Mouse IgG2a κ 100 μg 1 mg/mL

BACKGROUND: Homeobox genes (HOX gene), which encode DNA binding proteins that recognize specific sequences and modulate transcriptional activity, are expressed during embryogenesis with positional specificity. HOX-8 (Msx2) mRNA is highly expressed in immature tumors, including a yolk sac tomor, seminoma and choriocarcinoma, and that its expression is higher in various tumors of epitherial origin than in the corresponding normal tissues. Msx2 gene expression is frequently activated in carcinoma-derived cell lines. The gene is inactive in NIH/3T3 cells but is transcriptionally activated after transformation by v-Ki-ras, suggesting that the Msx2 may play a positive role in cell transformation.

SOURCE: This antibody was purified from hybridoma (clone 2E12) supernatant using protein A agarose. This hybridoma was established by fusion of mouse myeloma cell P3U1 with Balb/c mouse splenocyte immunized with recombinant N-terminal amino acids of human Msx2 (1-77 aa.).

FORMULATION: 100 μg IgG in 100 μL volume of PBS containing 50% glycerol, pH 7.2. No preservative is contained.

STORAGE: This antibody solution is stable for one year from the date of purchase when stored at -20°C.

REACTIVITY: This antibody reacts with Msx2 on Western blotting.

APPLICATIONS:

Western blotting; 1 μg/mL for chemiluminescence

detection system

Immunoprecipitation; Not tested Immunohistochemistry; 10 µg/mL Immunocytochemistry; Not tested Flow cytometry; Not tested

Detailed procedure is provided in the following **PROTOCOLS**.

SPECIES CROSS REACTIVITY:

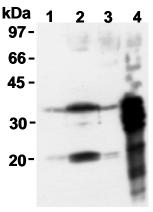
Species	Human	Mouse	Rat
Cells	HeLa, ZR-75-1, KB	Not Tested	Not Tested
Reactivity on WB	+		

INTENDED USE:

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REFERENCES:

- 1) Takahashi, C., et al., Oncogene 12, 2137-2146 (1996)
- 2) Suzuki, M., et al., Biochem. Biophys. Res. Commun. 194, 187-193 (1993)
- 3) Simeone, A., et al., Nature 320, 763-765 (1986)



Western blot analysis of Msx2 expression in HeLa (1), ZR-75-1 (2), KB (3) and recombinant (4) using M027-3.

PROTOCOLS:

SDS-PAGE & Western Blotting

- 1) Wash the cells 3 times with PBS and suspend with 10 volume of cold Lysis buffer (50 mM Tris-HCl, pH 7.2, 250 mM NaCl, 0.1% NP-40, 2 mM EDTA, 10% glycerol) containing appropriate protease inhibitors. Incubate it at 4 °C with rotating for 30 minutes, then sonicate briefly (up to 10 seconds).
- 2) Centrifuge the tube at 12,000 x g for 10 minutes at 4 °C and transfer the supernatant to another tube. Measure the protein concentration of the supernatant and add the cold Lysis buffer to make 8 mg/mL solution.
- 3) Mix the sample with equal volume of Laemmli's sample buffer.
- 4) Boil the samples for 3 minutes and centrifuge. Load 10 μL of the sample per lane in a 1 mm thick SDS-polyacrylamide gel for electrophoresis.
- 5) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm² for 1 hour in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% MeOH). See the manufacture's manual for precise

- transfer procedure.
- 6) To reduce nonspecific binding, soak the membrane in 10% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature, or overnight at 4 °C.
- 7) Incubate the membrane with primary antibody diluted with PBS, pH 7.2 containing 1% skimmed milk as suggest in the **APPLICATIONS** for 1 hour at room temperature. (The concentration of antibody to be used will be depend on condition.)
- 8) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 minutes x 3 times).
- 9) Incubate the membrane with the 1:10,000 HRP-conjugated anti-mouse IgG (MBL; code no. 330) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature.
- 10) Wash the membrane with PBS-T (10 minutes x 3 times).
- 11) Wipe excess buffer on the membrane, then incubate it with appropriate chemiluminescence reagent for 1 minute.
- 12) Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.
- 13) Expose to an X-ray film in a dark room for 3 minutes.
- 14) Develop the film as usual. The condition for exposure and development may vary.

(Positive controls for western blotting; HeLa, ZR-75-1, KB, recombinant)

<u>Immunohistochemical staining for paraffin-embedded sections: SAB method</u>

- 1) Deparaffinize the sections with Xylene 3 times for 3-5 minutes each.
- 2) Wash the slides with Ethanol 3 times for 3-5 minutes each.
- 3) Wash the slides with PBS 3 times for 3-5 minutes each.
- 4) Remove the slides from PBS and cover each section with 3% H₂O₂ for 10 minutes at room temperature to block endogenous peroxidase activity. Wash 3 times in PBS for 5 minutes each.
- 5) Remove the slides from PBS, wipe gently around each section and cover tissues with Protein Blocking Agent (Ultratech HRP Kit; IMMUNOTECH, code no. IM-2391) for 5 minutes to block non-specific antibody staining. Do not wash.
- 6) Tip off the blocking buffer, wipe gently around each section and cover tissues with primary antibody diluted with PBS containing 1% BSA as suggest in the **APPLICATIONS**.
- 7) Incubate the sections for 1 hour at room temperature.
- 8) Wash the slides 3 times in PBS for 5 minutes each.
- 9) Wipe gently around each section and cover tissues with Polyvalent Biotinylated Antibody (Ultratech HRP Kit). Incubate for 10 minutes at room temperature. Wash as in step 8.
- 10) Wipe gently around each section and cover tissues with Streptavidin-Peroxidase (Ultratech HRP Kit). Incubate for 10 minutes at room temperature. Wash as in step 8.
- 11) Visualize by reacting for 10-20 minutes with substrate solution containing 7.5 mg DAB, 40 μ L of 30% H₂O₂ in 150 mL PBS. *DAB is a suspect carcinogen and must be handled with care. Always wear gloves.

- 12) Wash the slides in water for 5 minutes.
- 13) Counter stain in hematoxylin for 1 minute, wash the slides 3 times in water for 5 minutes each, and then immerse the slides in PBS for 5 minutes. Dehydrate by immersing in Ethanol 3 times for 3 minutes each, followed by immersing in Xylene 3 times for 3 minutes each.
- 14) Now ready for mounting.