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Fluorescent protein-protein interaction visualization

# Fluoppi<sup>†</sup>

PPIs Detection Reagent : Fluoppi [ p53-MDM4 ]

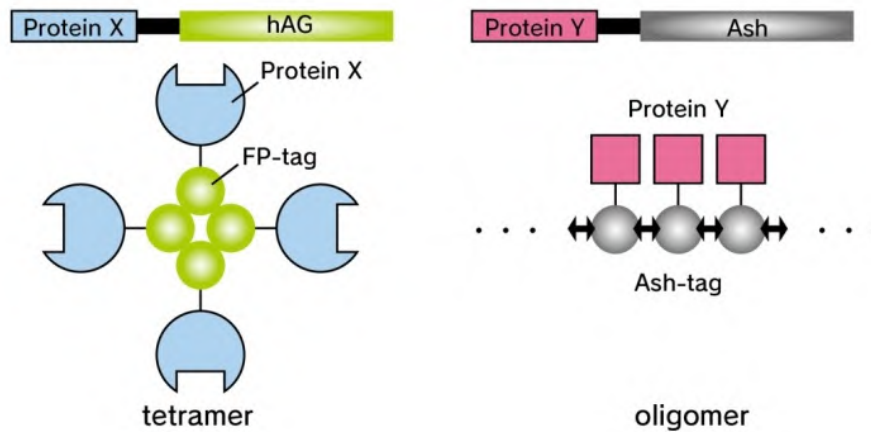
Code: AM-P0001

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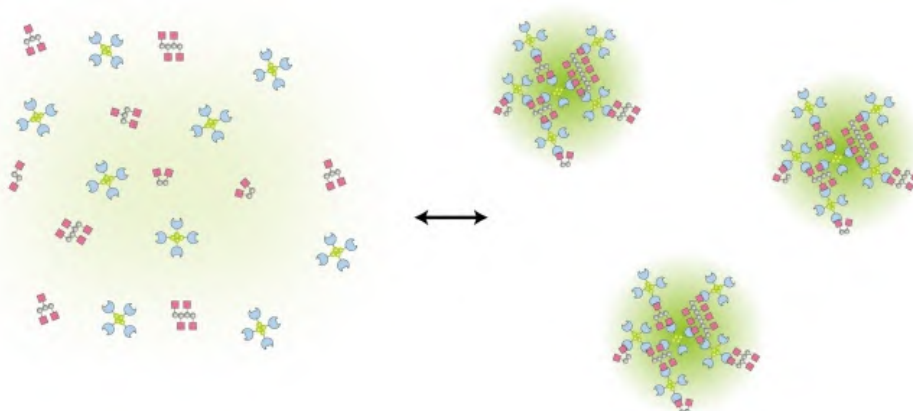
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## 1. Introduction

**Fluoppi** is a technology providing an easy way to visualize protein-protein interactions (PPIs) with a high signal to noise ratio. It employs an oligomeric assembly helper tag (Ash-tag) and a tetrameric fluorescent protein tag (FP-tag) to create detectable fluorescent puncta when there are interactions between two proteins fused to the tags. Schematic images are illustrated in Figure 1, where genetic fusion of protein X with FP-tag, and Y with Ash-tag creates a tetrameric fluorescent fusion protein X-FP and an oligomeric fusion protein Y-Ash respectively. Because each fusion protein has multiple Xs or Ys, the interaction between X and Y causes phase-separated droplets where the fluorescence by X-FP is concentrated and detectable as fluorescent puncta (Fig. 2).



**Figure 1** | Key components of Fluoppi technology



**Figure 2** | Mechanism of action

## 2. Fluoppi : Ash-hAG [ p53-MDM4 ]

This product contains two expression cassettes for detecting p53-MDM4 interaction in living cells. One encodes a fusion protein Ash/p53, and the other encodes humanized Azami-Green (hAG)/MDM4. MDM4 is also known as MDMX. Partial sequences responsible for this interaction are used for this product. Co-transfection of DNA cassettes, Ash/p53 and hAG/MDM4, results in formation of cytoplasmic fluorescent puncta. After addition of p53-MDM4 PPI inhibitors, the puncta disappeared within a day, indicating the p53-MDM4 complex was disrupted.

## 3. Product Components and Storage Condition

DNA cassettes	Amount:	Form
Ash/p53	10 µg	Dry form
hAG/MDM4	10 µg	Dry form

Reconstitute in 10-50 µL of sterilized distilled water before use.

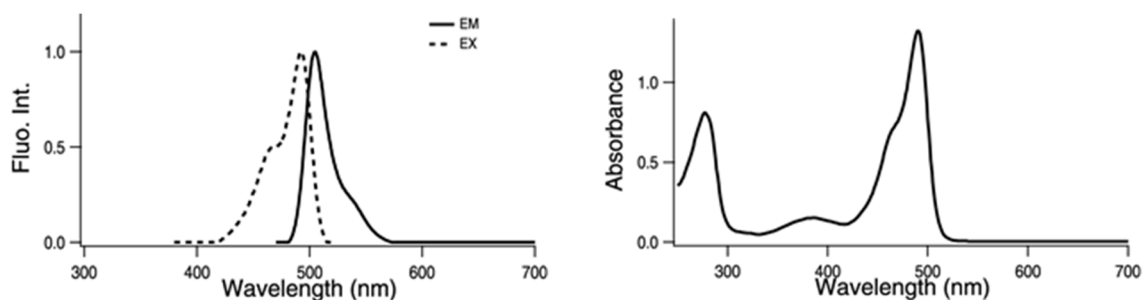
Storage condition: Store at -20°C. Reconstituted solution should be kept at -20°C.

## 4. Additional Materials Required

- Cell culture related materials (Mammalian cells, Cell culture medium, Cell culture dish, etc.)
- Transfection reagents or equipment.
- Buffer for imaging (HBSS, PBS, Good's Buffers, etc.)
- Fluorometric detector (Fluorescence microscopy or Plate imager)

## 5. Properties of Fluorescent protein “Azami-Green”

humanized Azami-Green (hAG), cloned from the stony coral (Azami-sango in Japanese), absorbs light maximally at 492 nm and emits green light at 505 nm. hAG forms tetramer and is featured by its fast maturation and highly photo and pH stable nature. The gene codon is optimized for mammalian cells.



Fluorescent protein	Excitation/Emission maximum (nm)	Extinction coefficient ( $M^{-1}cm^{-1}$ )	Fluorescence quantum yield	pKa
hAG	492/505	72,300 (492 nm)	0.67	<5.0

## 6. Expression Cassettes

Both open reading frames are driven by the CMV promoter in mammalian cells.



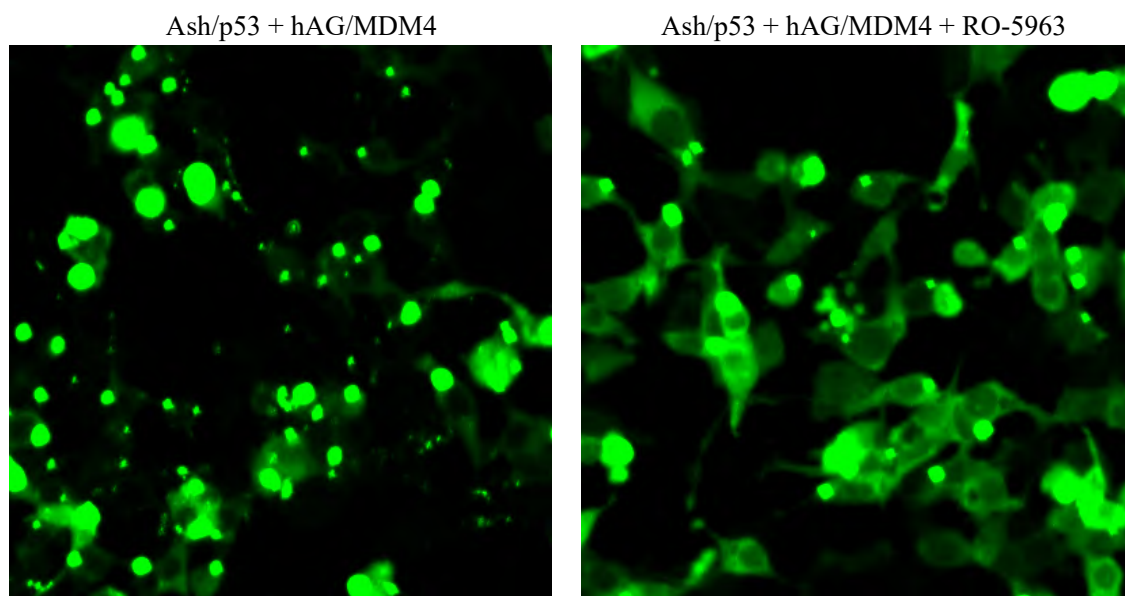
## 7. Example of Procedure

### [Transfection]

HEK293 cells were grown in DMEM (Sigma; code No. D5796) supplemented with 10% fetal bovine serum (FBS) and 1% Pen Strep (Gibco; code No. 15140-122) at 37°C in 5% CO<sub>2</sub> atmosphere. 4x10<sup>6</sup> cells were transfected with this product (4 µg each) using the Neon<sup>®</sup> Transfection System (Thermo Fisher Scientific) with the following parameters, 1000 V, 40 ms, 1 pulse. Cells were seeded in 96-well plates (BD Biosciences; code No.354640) at 4x10<sup>4</sup> cells per well with 100 µL medium. After incubation for 20-24 hours followed by replacing the growth medium with imaging buffer (20 mM HEPES-NaOH pH 7.4/HBSS) , cells were subjected to analysis.

### [Imaging]

A wide field fluorescence microscopy was used to observe PPI. Excitation of hAG fluorescence was performed by a 75-W Xenon lamp with a BP460-480HQ filter (Olympus). Emitted light was detected by an ORCA-Flash4.0 sCMOS camera (Hamamatsu Photonics) with a BA495-540HQ band pass filter (Olympus) and a 485 nm dichroic mirror (Olympus). MetaMorph software (Molecular Devices) was used for data collections and analysis.



**Figure 3** | HEK293 cells transiently expressing both Ash/p53 and hAG/MDM4 were observed at 24 hours after addition of 0.01 µM (left) and 25 µM (right) RO-5963\*. The interactions were observed as fluorescent puncta (left), and disruptions of the PPI by RO-5963 resulted in cytoplasmic diffused distributions of fluorescence (right).

\*An inhibitor of p53-MDM4 binding (Graves, B., *et al.* 2012).

8. References

Watanabe T, *et al.*, Genetic visualization of protein interactions harnessing liquid phase transitions. Sci Rep. 7, Article number: 46380 (2017) [PMID: 28406179 ]

Graves, B., *et al.*, Activation of the p53 pathway by small-molecule-induced MDM2 and MDMX dimerization. Proc. Natl. Acad. Sci. USA. 109, 11788-11793. (2012) [PMID: 22745160]

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