

# Anti-AAV8 Antibody ELISA Kit for Monkey

Code No. 5183/5184

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## Intended Use

**Anti-AAV8 Antibody ELISA Kit for Monkey** can be used to measure monkey anti-AAV8 antibodies in sera.

To measure monkey anti-AAV8 antibody, please use two Anti-AAV8 Antibody ELISA Kits (MBL; Code No. 5183 and 5184) in combination. The two kits are sufficient to produce one 96-well microplate.

Individual users should determine appropriate conditions when using other types of samples.

**This assay kit is for research use only and not for use in diagnostic or therapeutic procedures.**

## Storage

- Upon receipt store #5183 components at -80°C, and #5184 components at 2-8°C.
- Don't expose reagents to excessive light.

## Materials Provided

**Code No. 5183** (store at -80°C)

| Name                               | Materials                                 | Quantity        |
|------------------------------------|---|-----------------|
| <b>AAV8 empty capsid</b>           | Antigen for solid phase to the Microplate | 20 µL × 1 vial  |
| <b>Anti-AAV8 antibody standard</b> | anti-AAV8 antibodies                      | 200 µL × 1 vial |

\* The components should be stored at -80°C. Avoid repeated freeze-thaw cycles.

**Code No. 5184** (store at 2-8°C)

| Name                           | Materials  | Quantity           |
|--------------------------------|--|--------------------|
| <b>Microplate</b>              | Microwell strips   | 8-well × 12 strips |
| <b>Coating buffer</b>          | Buffer for coating microwells with AAV8 empty capsid                 | 20 mL × 1 bottle   |
| <b>Blocking buffer</b>         | Buffer for blocking microwells (Ready-to-use)                        | 100 mL × 1 bottle  |
| <b>Sample diluent</b>          | Buffer for diluting samples (Ready-to-use)                           | 30 mL × 1 bottle   |
| <b>Conjugate diluent</b>       | Buffer for diluting HRP conjugated antibody (Ready-to-use)           | 14 mL × 1 bottle   |
| <b>HRP conjugated antibody</b> | HRP conjugated anti-IgG (Monkey) polyclonal antibody (100x)          | 150 µL × 1 vial    |
| <b>Wash concentrate (20x)</b>  | Buffer for washing microwells (20x)                                  | 50 mL × 1 bottle   |
| <b>Substrate reagent</b>       | TMB/H <sub>2</sub> O <sub>2</sub> solution (Ready-to-use)            | 20 mL × 1 bottle   |
| <b>Stop solution</b>           | 0.5N H <sub>2</sub> SO <sub>4</sub> solution (0.25 M) (Ready-to-use) | 20 mL × 1 bottle   |
| <b>Plate seals</b>             | Plate seals  | 3 pieces           |

## Materials Required but not Provided

- **Pipettors (single and multichannel):** 2-20 µL, 20-200 µL and 200-1,000 µL precision pipettors with disposable tips.
- **Precision repeating pipettor**
- **Plastic tubes** (1.5 mL, 15 mL etc.)
- **(Optional) Microplate washer:** Manual washing is possible.
- **Reagent reservoirs**
- **Deionized water of the highest quality**
- **Disposable paper towels**
- **Plate reader:** capable of measuring absorbance in 96-well plates at dual wavelengths of 450 nm/620 nm. The plate can also be read at a single wavelength of 450 nm, which will give a somewhat higher reading.
- **(Optional) Software package facilitating data generation and analysis**

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## Precautions and Recommendations

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- All reagents need to be brought to room temperature (18-27°C) before use.
- Do not use kit components beyond the indicated kit expiration date.
- Do not mix reagents with different batches and kits.
- Do not mouth pipette or ingest any of the reagents.
- Fresh samples should be used. Aliquot each sample and store below -20°C if necessary. Avoid repeated freezing and thawing. Never store the samples at 4°C, as samples might be affected by storage at this temperature.
- AAV8 empty capsid is easy to adsorb on polystyrene. When handling AAV8 empty capsid, please use polypropylene tubes and tips.
- The buffers and reagents in this kit may contain preservatives. Care should be taken to avoid direct contact with these reagents. During disposal, flush with plenty of water and/or handle it according to the regulations of the facility.
- Dispose tetra-methylbenzidine (TMB) containing solutions in compliance with local regulations.
- Avoid contact with the acidic **Stop solution** and **Substrate reagent**, which contains hydrogen peroxide. Protect eyes and skin and handle with care. In case of contact with the **Stop solution** and the **Substrate reagent**, wash skin thoroughly with water and seek medical attention, when necessary.
- Wear gloves and eye protection when handling immunoassay materials and samples of human/monkey origin and these reagents.
- **Biological samples may be contaminated with infectious agents. Do not ingest, expose to open wounds, or breathe aerosols. Wear protective gloves and dispose of biological samples properly.**
- **AAV8 empty capsid is a kit component subject to the regulations of the Cartagena, Law Concerning the Conservation and Sustainable Use of Biological Diversity through Regulations on the Use of Living Modified Organisms. Please handle it carefully according to the regulations of each user's facility.**
- **AAV8 empty capsid is known as non-infectious against human, however, instruments used in this assay should be treated according to the regulations of each facility after the assay.**  
(e.g.)
  - Soak in 2% glutaraldehyde solution (final concentration) for more than one hour.
  - Soak in 0.1% sodium hypochlorite solution (available chloric: approximately 1,000 ppm.) for more than one hour.
  - Autoclave at 121°C for more than 20 minutes.

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## Sample Collection and Storage

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**Serum:** Use a serum separation tube and separate the serum according to the manufacturer's manual.

Fresh samples should be used. Aliquots of serum may also be stored at below -20°C for extended periods of time. Avoid repeated freeze-thaw cycles.

**Other biological samples:** Not validated by MBL.

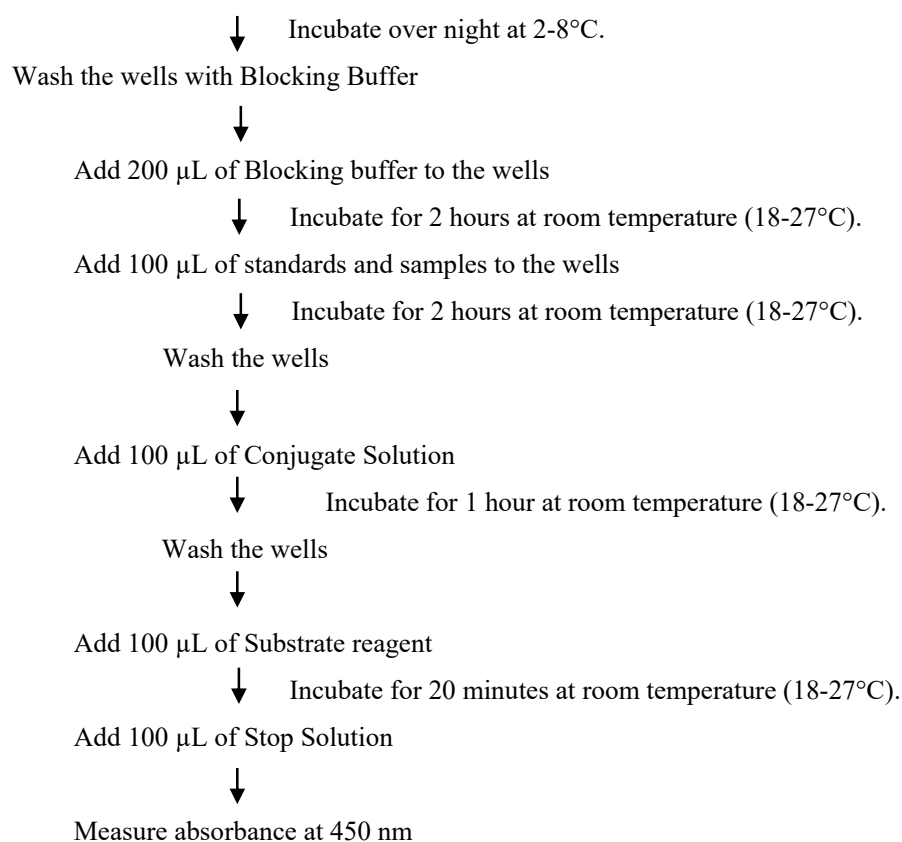
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## Summary of Procedure

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To prepare the antigen-**coated** wells: Add 100 µL of AAV8 Antigen Solution to the wells

To prepare the antigen-**UNCOATED** wells: Add 100 µL of Coating Buffer to the wells



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## Detailed Protocol

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The **Anti-AAV8 Antibody ELISA Kit** is provided with removable strips of wells so the assay can be carried out on separate occasions. Experimental conditions may vary. To measure monkey anti-AAV8 antibody, please use two Anti-AAV8 Antibody ELISA Kits for Monkey (MBL; Code No. 5183 and 5184) in combination. The two kits are sufficient to produce one 96-well microplate. All samples and the positive control should be assayed in duplicate. Disposable pipette tips and reagent troughs should be used for all liquid transfers to avoid cross-contamination of reagents or samples.

### <Preparation of Working Solutions>

All reagents need to be brought to room temperature prior to assay use. **Sample diluent** should be inverted and mixed before use.

#### 1. Wash Buffer

Prepare "Wash Buffer" by adding 50 mL of the **Wash concentrate (20x)** to 950 mL of deionized (distilled) water (ddH<sub>2</sub>O). Mix well.

#### 2. AAV8 Antigen Solution

The "AAV8 Antigen Solution" should be used immediately after preparation. Prepare "AAV8 Antigen Solution" by diluting **AAV8 empty capsid** with **Coating Buffer** according to the attached lot specific document "*Preparation of Antigen*". **AAV8 empty capsid** should be thawed at room temperature (18-27°C), and mixed gently. Do not vortex strongly.

**\*AAV8 empty capsid is easy to adsorb on polystyrene. When handle AAV8 empty capsid, please use polypropylene tubes and tips.**

#### 3. Anti-AAV8 Antibody Standard Solution (Master Standard)

Prepare "Anti-AAV8 Antibody Standard Solution" by diluting **Anti-AAV8 antibody standard, 25-fold** with **Sample diluent** by gently mixing (*e. g.*, add 40 µL of **Anti-AAV8 antibody standard** to 960 µL of **Sample diluent**). After thawing **Anti-AAV8 antibody standard**, dispense it in small aliquots (*e.g.* 50 µL) to plastic micro-centrifuge tubes and store below -80°C to avoid non-specific adsorption to glass surface and multiple freeze-thaw cycles.

The prepared Anti-AAV8 Antibody Standard Solution is referred to as the Master Standard.

\* Sample diluent should be inverted and mixed before use.

Prepare Standard dilution series as follows:

Use the Master Standard to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The Master Standard (Std.1) serves as the highest standard.

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|        | Volume of Standard               | Sample diluent | Dilution ratio    |
|--------|----------------------------------|----------------|-------------------|
| Std. 1 | 1,000 $\mu$ L of Master Standard | 0 $\mu$ L      | x25 (100 U/mL)    |
| Std. 2 | 500 $\mu$ L of Std. 1            | 500 $\mu$ L    | x50 (50 U/mL)     |
| Std. 3 | 500 $\mu$ L of Std. 2            | 500 $\mu$ L    | x100 (25 U/mL)    |
| Std. 4 | 500 $\mu$ L of Std. 3            | 500 $\mu$ L    | x200 (12.5 U/mL)  |
| Std. 5 | 500 $\mu$ L of Std. 4            | 500 $\mu$ L    | x400 (6.25 U/mL)  |
| Std. 6 | 500 $\mu$ L of Std. 5            | 500 $\mu$ L    | x800 (3.125 U/mL) |

**Note:** Do not use a Repeating pipette. Change tips for every dilution. Wet tip with Sample diluent before dispensing. The Sample diluent serves as the Blank.

4. Conjugate Solution

Prepare "Conjugate Solution" by diluting **HRP conjugated antibody, 100-fold** with **Conjugate diluent**. Prepare only a sufficient amount of the Conjugate Solution immediately before use (*e. g.*, add 100  $\mu$ L of **HRP conjugated antibody** to 9,900  $\mu$ L of **Conjugate diluent**).

5. Other reagents are ready-to-use.

**<Preparation of samples>**

Samples require the proper dilution ratio if necessary. As an example, serum samples used in MBL were diluted 128-fold with **Sample diluent**. If the measured value falls outside the range of the calibration curve, we recommend verifying the dilution ratio and retesting. (*e. g.*, add 10  $\mu$ L of each sample to 1,270  $\mu$ L of **Sample diluent**)

\* Sample diluent should be inverted and mixed before use.

## <Assay Procedure>

### Coating

\*Please note that both antigen-coated wells and antigen-UNCOATED wells should be prepared for the assay.

1. To prepare the antigen-coated wells, pipette **100 µL** of AAV8 Antigen Solution to the wells of **Microplate** with a multichannel pipet.
2. To prepare the antigen-UNCOATED wells, pipette **100 µL** of Coating buffer to the wells of **Microplate** with a multichannel pipet.
3. Incubate the plate overnight at 2~8°C. Please be careful not to evaporate the solution in the wells.
4. Discard the solution in the wells, and wash the plate twice by filling each well with **Blocking Buffer** (200 µL) using a multi-channel pipette. Tap the plate on a paper towel to remove any remaining solution.

\*Wash as quickly as possible, do NOT let wells dry up.

An example of using Microwell strips;

| Antigen-coated wells |       |       |       | Antigen-UNCOATED wells |    |    |    |     |     |     |     |
|----------------------|-------|-------|-------|------------------------|----|----|----|-----|-----|-----|-----|
| Std.1                | Std.1 | Std.1 | Std.1 | S2                     | S2 | S2 | S2 | S10 | S10 | S10 | S10 |
| Std.2                | Std.2 | Std.2 | Std.2 | S3                     | S3 | S3 | S3 | S11 | S11 | S11 | S11 |
| Std.3                | Std.3 | Std.3 | Std.3 | S4                     | S4 | S4 | S4 | S12 | S12 | S12 | S12 |
| Std.4                | Std.4 | Std.4 | Std.4 | S5                     | S5 | S5 | S5 | S13 | S13 | S13 | S13 |
| Std.5                | Std.5 | Std.5 | Std.5 | S6                     | S6 | S6 | S6 | S14 | S14 | S14 | S14 |
| Std.6                | Std.6 | Std.6 | Std.6 | S7                     | S7 | S7 | S7 | S15 | S15 | S15 | S15 |
| Blank                | Blank | Blank | Blank | S8                     | S8 | S8 | S8 | S16 | S16 | S16 | S16 |
| S1                   | S1    | S1    | S1    | S9                     | S9 | S9 | S9 | S17 | S17 | S17 | S17 |

### Blocking

5. Pipette **200 µL** of **Blocking buffer** to the wells with a multichannel pipet.
6. Incubate the plate for 2 hours at room temperature (18-27°C).
7. Discard the solution in the wells, and tap the plate on a paper towel to remove any remaining solution.

### 1<sup>st</sup> Reaction

8. Pipette **100 µL** of prepared Standards and diluted samples to the wells, both antigen-coated wells and antigen-UNCOATED wells respectively.

9. Incubate the plate for 2 hours at room temperature (18-27°C).

#### Washing

10. Discard the solution in the wells.

11. Wash the plate 4 times by filling each well with Wash Buffer using a squirt bottle, multi-channel pipette, manifold dispenser or microplate washer. Tap the plate on a paper towel to remove any remaining solution.

\* When using an auto-washer, appropriate washing times may vary depending on the instrument.

#### 2<sup>nd</sup> Reaction

12. Pipette **100 µL** of Conjugate Solution to the wells with a multichannel pipet.

13. Incubate the plate for 1 hour at room temperature (18-27°C).

#### Washing

14. Discard the solution in the wells.

15. Wash the plate 4 times as in step 11.

#### Enzyme Reaction

16. Pipette **100 µL** of **Substrate reagent** to the wells with a multichannel pipet.

17. Incubate the plate for 20 minutes at room temperature (18-27°C).

#### Reading

18. Pipette **100 µL** of **Stop solution** to the wells with a multichannel pipet.

19. Measure absorbance in each well using a spectrophotometric microplate reader at dual wavelengths of 450/620 nm. The primary wavelength is 450 nm, and the reference wavelength is 620 nm. Read the microplate at 450 nm if only a single wavelength can be used. Wells must be read within 30 minutes of adding the Stop solution.

**Note-1:** Each solution and reagent should be used at room temperature (18-27°C).

**Note-2:** Use new disposable tips, reservoirs and paper towels at each step to avoid contamination.

**Note-3:** Once the solution has been poured into the reservoir, never return it to the bottle.

**Note-4:** Complete removal of liquid at each step is essential to good performance.

**Note-5:** Ensure that the back of the plate is clean and dry, and that no air bubbles are present on the surface of the liquid in the wells before reading.

**Note-6:** The incubation time for color development may vary depending on the environment such as temperature. It can be adjusted according to the coloring intensity.



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## Calculations

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Anti-AAV8 antibody concentration in each sample can be calculated using a 4-parameter logistic regression equation.

- 1) Calculate SPEC O.D. for each well  
(O.D. value of antigen-coated wells) – (O.D. value of antigen-UNCOATED wells) = SPEC O.D.
- 2) Plot the SPEC O.D. values average of the duplicate measurements for each standard except Blank wells (Y) versus the known concentration (X) of each standard, and draw the best fit curve. Most microtiter plate readers perform automatic calculations of analyte concentration. At the point of intersection, extend a vertical line to the x-axis and read the corresponding concentration (U/mL).

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## Measurement Range

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The measurement range is 3.1 U/mL to 100 U/mL. Any sample reading higher than the highest standard should be diluted with Sample diluent in higher dilution and re-assayed. Dilution factors need to be taken into consideration in calculating the concentration of the sample.

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## Quality Control

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To assure the validity of the results, each assay must include both Blank and the highest standard. The net O.D. values (450 nm) of these controls must fall within the ranges listed below. If O.D. values do not meet the requirements, the assay is invalid and requires repeat of the assay.

- O.D. values of Blank  $\leq 0.09$   
SPEC O.D. values of the highest standard (Std. 1)  $\geq 1.0$

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## Troubleshooting

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1. All samples and Blank should be assayed in duplicate, using the protocol described in the **Detailed Protocol**. Incubation times or temperatures significantly different from those specified may give erroneous results.
2. Do not dry the plate as it can adversely affect the assay results. Immediately add each solution step by step.

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## Reagent Stability

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#5183 components must be stored at -80°C, and #5184 components must be stored at 2-8°C. Reagents should not be used beyond the stated expiration date.

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## Assay Characteristics

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### 1. Precision

#### Intra-assay Precision (Precision within an assay)

Samples of known concentration were tested in septuplicate on one plate to assess intra-assay precision.

- Intra-assay (Within-Run, n=6)

|        | Sample 1 | Sample 2 | Sample 3 |
|--------|----------|----------|----------|
| 1      | 64.5     | 32.3     | 18.5     |
| 2      | 59.8     | 34.4     | 18.7     |
| 3      | 55.4     | 31.3     | 17.7     |
| 4      | 52.6     | 30.7     | 18.0     |
| 5      | 52.6     | 28.4     | 18.7     |
| 6      | 55.6     | 30.7     | 18.4     |
| Max.   | 64.5     | 34.4     | 18.7     |
| Min.   | 52.6     | 28.4     | 17.7     |
| Mean   | 56.8     | 31.3     | 18.3     |
| SD     | 4.6      | 2.0      | 0.4      |
| CV (%) | 8.1      | 6.3      | 2.2      |

#### Inter-assay Precision (Precision between assays)

Three septuplicate-measurements were performed to assess inter-assay precision. Samples of known concentration were used.

- Inter-assay (Run-to-Run, n=3)

|        | Sample 1 | Sample 2 | Sample 3 |
|--------|----------|----------|----------|
| 1      | 56.8     | 31.3     | 18.3     |
| 2      | 61.6     | 32.7     | 15.4     |
| 3      | 77.3     | 36.2     | 17.1     |
| Mean   | 65.2     | 33.4     | 16.9     |
| SD     | 10.7     | 2.5      | 1.5      |
| CV (%) | 16.4     | 7.6      | 8.9      |

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## Examples of Test Results

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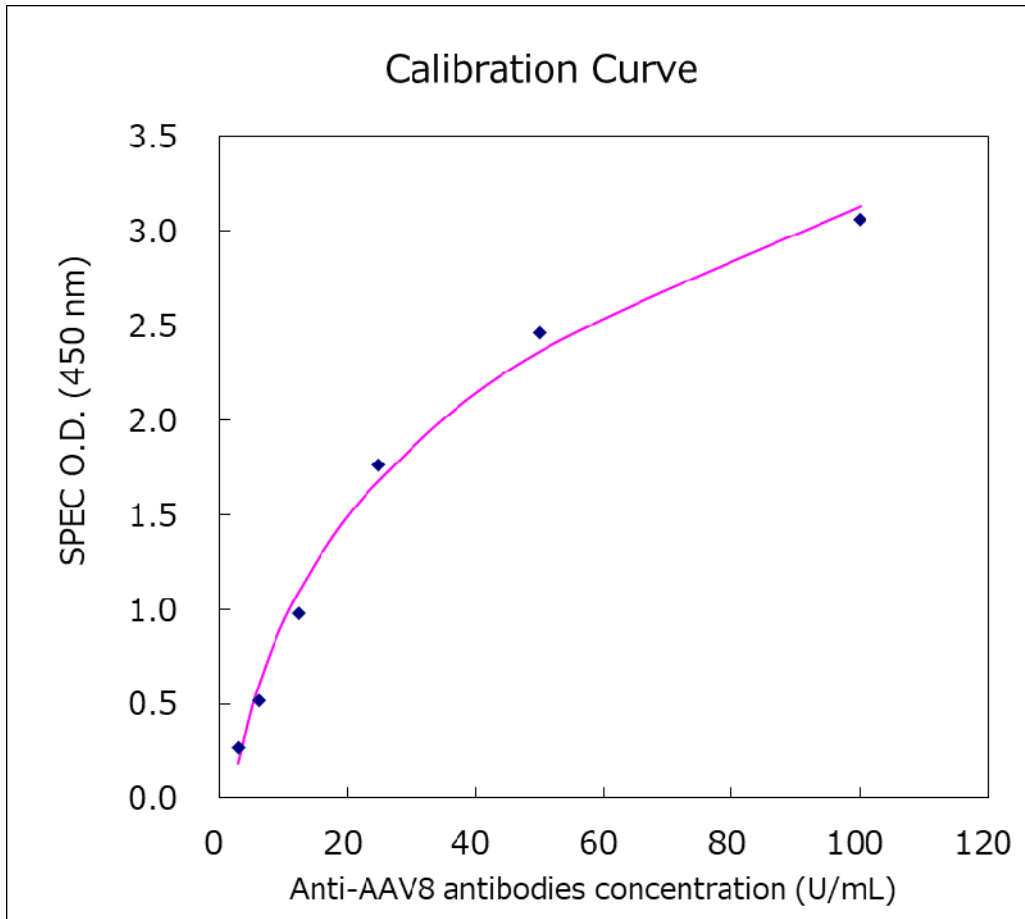


Fig.1 Anti-AAV8 antibodies calibration curve

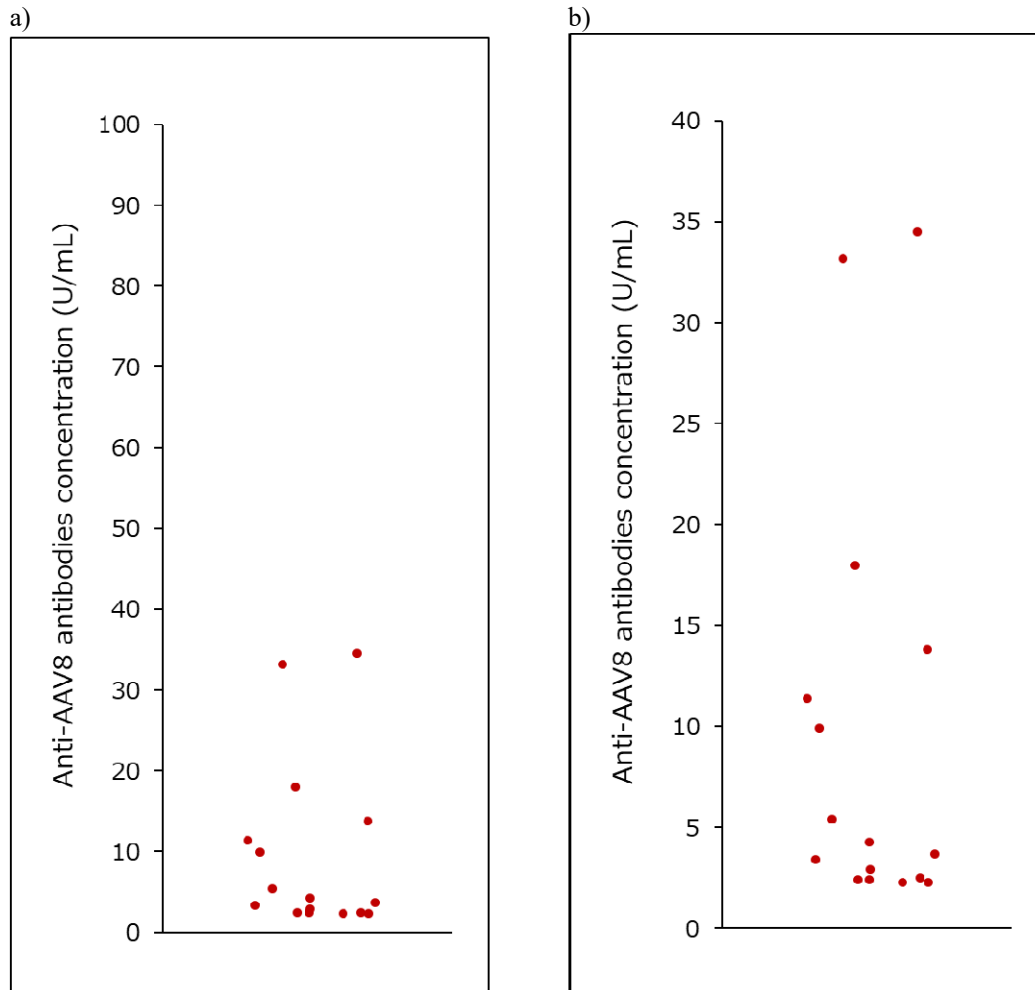


Fig.2 The Anti-AAV8 antibody concentration serum derived from Monkey (n=16). These samples were diluted 128-fold with Sample diluent.

- a) A figure showing distribution of measured values of 16 samples.  
b) An enlarged figure of the plot of less than 40 U/mL. (16 samples)

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## Preparation of Antigen

Lot No. XXXXXX

To prepare "AAV8 Antigen Solution" by diluting **AAV8 empty capsid** with Coating Buffer.

1. Thaw **AAV8 empty capsid** at room temperature (18-27°C).
2. Dilute the thawed **AAV8 empty capsid** X-fold with Coating Buffer.  
(e. g., Add the thawed Y  $\mu$ L of **AAV8 empty capsid** to Z  $\mu$ L of Coating Buffer)

\* **AAV8 empty capsid** should be mixed with Coating Buffer immediately. The "AAV8 Antigen Solution" should be used immediately after preparation.

\* AAV8 empty capsid is easy to adsorb on polystyrene. When handling AAV8 empty capsid, please use polypropylene tubes and tips.